

DOTTORATO DI RICERCA IN BIOLOGIA CELLULARE E DELLO SVILUPPO

40th CYCLE Project proposal for a Sapienza PhD scholarship

Main research line

Title: Exploring sensory-motor neuron innervation of skeletal muscle affected in Duchenne Muscular Dystrophy (DMD)

Supervisor: Maria Egle De Stefano; egle.destefano@uniroma1.it

Website: <https://corsidilaurea.uniroma1.it/it/users/egledestefanouniroma1it>

Co-Supervisor: Prof. Francesca Grassi, Dip. Fisiologia e Farmacologia "Vittorio Erspamer", Sapienza Universita di Roma

Summary

In Duchenne Muscular Dystrophy (DMD), the absence of Dp427 and one or more of its short isoforms has a significant impact not only on skeletal muscle. In the central nervous system, several molecular pathways are directly and/or indirectly affected by the lack of dystrophin, among which the inhibitory GABA signaling in specific brain regions. The spinal cord (SC), which is the portion of the central nervous system directly connected to the muscle via the double stream of sensory and motor neurons (MNs), is likely to endure the impact of dystrophin deficiency, along with signals associated with muscle atrophy. Nevertheless, the impact of DMD pathology on sensory neurons innervating skeletal muscles and their intraspinal connections with ventral MNs remains largely unexplored. A recent report suggests that proprioception, largely dependent on the activity of muscle spindles, might be impaired in DMD patients, although nociception and tactile sensitivity are preserved. Experimental evidence highlights anatomical and/or functional alterations in muscle spindles in both DMD patients and the mdx mouse model of the disease and modifications in intraspinal connections between sensory and MNs. GABA receptors play a major role in modulating the sensory-induced activation of MNs, and alterations in the abundance and composition of GABA receptors have been reported in mdx vs. wild type mice. Together, these abnormalities in the sensory-motor connections might compromise muscle reflexes, strength, and coordination, exacerbating motor imbalances and muscular deficits associated with the disease. This project seeks to elucidate the extent to which structural and functional interplay between skeletal muscles, sensory neurons, MNs, and intraspinal sensory-motor connections is affected in DMD. Utilizing the mdx mouse and its wild-type control, the study will employ various approaches and techniques, to perform a comprehensive analysis of morphological and functional alterations in sensory ganglia (neurons and satellite cells), MNs, and intraspinal connectivity. The hypothesis that muscle necrosis has repercussions on sensory and MNs, possibly through the activation of glial cells will be explored by treating mice with TRAM-34, a selective blocker of ion channels expressed in macrophages/microglia, ganglionic satellite cells and fibroblasts, which reduces muscle fibrosis and promotes an anti-inflammatory profile of muscle macrophages and spinal microglia. The ultimate objective is to provide insights into potential innovative therapeutic approaches focused not only on improving the muscular condition but also on enhancing

muscle-SC sensory-motor interaction.

Pertinent Publications of the proponent (last 5 years)

1. Botticelli E, Guerriero C, Fucile S, **De Stefano ME**, Matera C, Dallanoce C, De Amici M, Tata AM (2023) Activation of $\alpha 7$ nicotinic acetylcholine receptors via metabotropic downstream signaling pathways may improve Schwann cell regeneration potential. *Cells* 12:1494. doi: 10.3390/cells12111494; IF: 7.666; 5-year IF: 7.677
2. Rosito M, Sanchini C, Gosti G, Moreno M, De Panfilis S, Giubettini M, Debillis D, Catalano F, Peruzzi G, Marotta R, Indrieri A, De Leonibus E, **De Stefano ME**, Ragazzino D, Ruocco G, Di Angelantonio S, Bartolini (2023) Microglia reactivity entails microtubule remodeling from acentrosomal arrays. *Cell Rep* 42:112104. doi: 10.1016/j.celrep.2023.112104; IF: 9.995; 5-year IF: 10.990
3. Morotti M, Garofalo S, Cocozza G, Antonangeli F, Bianconi V, Mozzetta C, De Stefano ME, Capitani R, Wulff H, Limatola C, Catalano M, F Grassi (2022) Muscle damage in dystrophic *mdx* mice is influenced by the activity of Ca_{2+} -activated $\text{KCa}_{3.1}$ channels. *Life* 12:538. doi: 10.3390/life12040538
4. **De Stefano ME**, Ferretti V, Mozzetta C (2021) Synaptic alterations as a neurodevelopmental trait of Duchenne muscular dystrophy. *Neurobiol Dis* 168:105718. doi: 10.1016/j.nbd.2022.105718
5. Briatore F, Pregno G, Di Angelantonio S, Frola E, **De Stefano ME**, Vaillend C, Sassoë-Pognetto M, Patrizi A (2020) Dystroglycan mediates clustering of essential GABAergic components in cerebellar Purkinje cells. *Front Mol Neurosci* 13: 164. doi: 10.3389/fnmol.2020.00164
6. Persiconi I, Cosmi F, Guadagno NA, Lupo G **De Stefano ME** (2020) Dystrophin is required for the proper timing in retinal histogenesis: a thorough investigation on the *mdx* mouse model of Duchenne Muscular Dystrophy. *Front Neurosci* 14:760. doi:10.3389/fnins.2020.00760
7. Fragapane P, Cosmi F, De Stefano ME (2020) Cultured hippocampal neurons of dystrophic *mdx* mice respond differently from those of wild type mice to an acute treatment with corticosterone. *Exp Cell Res* 386:111715. doi:10.1016/j.yexcr.2019.111715

REFERENCES

1. Doorenweerd N (2020) Neuromuscul Disord 30:437-442. doi:10.1016/j.nmd.2020.05.001
2. Waite A et al. (2012) Trends Neurosci. 35: 487-496. doi:10.1016/j.tins.2012.04.004
3. De Stefano ME et al. (2022) Neurobiol Dis 168:105718. doi:10.3390/life12040538
4. Houzelstein D et al. (1992) J Cell Biol 119:811-21. doi:10.1083/jcb.119.4.811
5. Hoffman EP et al. (1988) Neuron 1:411-20. doi:10.1016/0896-6273(88)90191-2
6. Lidov HG (1996) Brain Pathol 6:63-77. doi:10.1111/j.1750-3639.1996.tb00783.x
7. Zarrouki F et al. (2022) Int J Mol Sci 23:12617. doi:10.3390/ijms232012617
8. Jerusalem F et al. (1974) Brain 97:123-30. doi:10.1093/brain/97.1.123
9. Theroux MC et al. (2008) Paediatr Anaesth 18:256-9. doi:10.1111/j.1460-9592.2008.02411.x
10. Ng SY, Ljubicic V (2020) EBioMedicine 61:103032. doi:10.1016/j.ebiom.2020.103032
11. Verma S et al. (2022) Mol Neurobiol 59:1502–1527. doi:10.1007/s12035-021-02658-6
12. Paredes-Redondo A et al. (2021) Sci Adv 7:eabi8787. doi:10.1126/sciadv.abi8787
13. Piotrkiewicz M et al. (1999 a) Clin Neurophysiol 110:1111-1122. doi:10.1016/s1388-2457(99)00051-6
14. Piotrkiewicz M et al. (1999 b) J Physiol (Paris) 93:167-173. doi:10.1016/s0928-4257(99)80147-8

15. Swash M, Fox KP (1976) J Neurol Sci 29:17-32. doi:10.1016/0022-510x(76)90077-0
16. Skuk D et al. (2010) Muscle Nerve 41:729-30. doi:10.1002/mus.21644
17. Troise D et al. (2014) Dev Med Child Neurol 56:881-7. doi:10.1111/dmcn.12469
18. Green D (2014) Dev Med Child Neurol 56:805-6. doi:10.1111/dmcn.12476
19. Prabhakar E, Lawson SN (1995) J Physiol 482:609-22. doi:10.1113/jphysiol.1995.sp020544
20. Mayeux V et al. (1996) Neuroscience 71:787-95. doi: 10.1016/0306-4522(95)00504-8
21. Gerwin L et al. (2020) J Physiol 598:1591-1609. doi: 10.1113/JP278563
22. Elbaz B et al. (2022) Cell Rep. 40:111130. doi: 10.1016/j.celrep.2022.111130
23. Hanani M, Spray D (2020) Nat Rev Neurosci 22:1-14. doi:10.1038/s41583-020-0333-z
24. Hanani M, Verkhratsky A (2021) Neurochem Res.46:2525-2537. doi:10.1007/s11064-021-03255-8
25. Haberberger RV et al. (2023) Cell Tissue Res 393:17-36. doi:10.1007/s00441-023-03770-w
26. Lu J et al. (2023) Stem Cell Rev Rep 19:358-367. doi:10.1007/s12015-022-10460-7
27. Bohlhalter S et al. (1996) J Neurosci 1996 16:283-97. doi:10.1523/JNEUROSCI.16-01-00283.1996
28. Rudomin P (2009) Exp Brain Res. 196:139–151. doi:10.1007/s00221-009-1758-9
29. Fink AJ, et al. (2014) Nature 509:43-8. doi: 10.1038/nature13276
30. Morotti M et al. (2022) Life12:538. doi:10.3390/life12040538
31. Onorato I et al. (2016) PLoS One 11:e0160950. doi:10.1371/journal.pone.0160950
32. Persiconi I et al (2020) *Front Neurosci* 14:760. doi:10.3389/fnins.2020.00760
33. Cocozza G et al (2018) Brain Behav Immun 73: 584-595. doi:10.1016/j.bbi.2018.07.002
34. De Stefano ME et al. (1997) Neuroscience 80:613–624. doi:10.1016/s0306-4522(97)00003-1
35. Fragapane P et al. (2020) *Exp Cell Res* 386:111715. doi:10.1016/j.yexcr.2019.111715
36. Carucci N et al. (2017) R. Soc Open Sci 4:160913. doi:10.1098/rsos.160913